First Biological Data for *Aspilodemon* Fischer (Hymenoptera: Braconidae: Hydrangeocolinae): Parasitoids of Cecidomyiid Fly Galls on Asteraceae in Brazil

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Abstract.—Biological data for the hydrangeocoline braconid genus *Aspilodemon* Fischer are reported for the first time. An *Aspilodemon* sp. from Brazil is reported as a parasitoid of three species of *Liodiplosis* spp. (Diptera: Cecidomyiidae) which induce galls on leaves of the liana, *Mikauia glomerata* (Asteraceae); it does not attack other gall-forming cecidomyiids on the same plant, and evidence suggests that it is ectoparasitic although direct observations of parasitoid larvae have not been made. How this affects our understanding of the evolution of gall formation and aphid parasitism by braconids is discussed.

The Hydrangeocolinae are a recently recognised subfamily of braconid wasps (Dowton et al. in press, Belshaw and Quicke in press), to date known from three genera, each with a southern distribution, viz Hydrangeocola Brèthes (= Kephalosema Fischer) from Chile, Aspilodemon Fischer, from Central and South America and Opiopterus Szépligeti (see Wharton 1993, Whitfield and Wharton 1997), from Australia. Until recently, some of these taxa were typically treated under the subfamily Opiinae (e.g. Fischer 1966, but see Wharton 1988) or more recently the Hormiinae, see e.g. Whitfield and Wharton (1997) or Rhyssalinae (Quicke et al. 1997) though several of these authors recognised, they do not fit well into any of these (see electronic appendix of Belshaw et al. 2000 for further discussion). Interest in the group was recently increased because molecular sequence data strongly indicate that they are not related to Opiinae, Hormiinae or Rhyssalinae, but instead form a sister group to the endemic Australian subfamily, Mesostoinae, and that together

these are the sister group of the well known Aphidiinae (Belshaw et al. 2000). This newly discovered relationship was particularly interesting as it appeared to suggest both a Gondwanan origin of the Aphidiinae despite their currently predominantly northern distribution, and a link with galls, because the most primitive extant aphids are gall formers, mesostoine braconids form galls on Banksia spp. (Proteaceae) (Austin and Dangerfield 1998) and the only hydrangeocoline for which some biology is known, Hydrangeocola, has been reared from unidentified galls on Hydrangea (Hydrangeaceae) (Brèthes 1927). Very little is known, however, about the biologies of the putatively most primitive extant aphidiines, and a direct association of any these with gall-forming aphids is vet to be confirmed.

Here we present the first rearing data for the genus *Aspilodemon* based on a probably undescribed species from Brazil. The species concerned is associated with cecidomyiid fly galls on *Mikania glomerata* Sprengel, and the available evidence

Locality	Morphological type of gall								
	Cylindrical	Spherical	Conical	Leat 'Vein	Buđ	Epidermis	Stem		
Parati	2070	1085	0	885	5	53	27		
Itatiaia	1385	468	0	574	11	17	22		
Ροςο	1061	583	17	272	5	17	0		
Picinguaba	861	471	188	305	2	38	9		
S. Órgãos	297	361	0	252	2	20	56		
Tijuca	0	6	824	51	0	141	0		

Table 1. Number of galls collected in each locality during study; gall types are illustrated in Gagné et al. (2001).

strongly suggests that it is a parasitoid of the fly larva. *M. glomerata* is a liana species belonging to the Asteraceae, and it occurs inside and on the edges of forests, flowering from August to December. The genus *Mikania* has 415 species mainly distributed in Central and South America, of which 171 species occur in Brazil (King and Robinson 1987). *M. glomerata* ranges from north-eastern Brazil down to the southern-most part of Brazil and just into Argentina and Paraguay (Ritter et al. 1992).

MATERIALS

Voucher specimens of the *Aspilodemon* sp. reared are deposited in each of the following collections: The Natural History Museum, London; the Entomological Collection of the Laboratório de Ecologia de Insetos, Department of Ecology, Universidade Federal do Rio de Janeiro. Morphological terminology follows Sharkey and Wharton (1997). The D2-D3 28S rDNA gene sequence for the species of *Aspilodemon* referred to here is deposited in EMBL: accession number AJ245685 (and has been used in phylogenetic analyses of Belshaw et al. 2000 and Quicke and Belshaw 1999).

Study areas and sampling.—The research was based in the Atlantic coast (Mata Atlântica) forests of Brazil which has one of the highest levels of biological diversity in the world, and is representative of humid tropical forests and their associated ecosystems (Mori et al. 1981). The localities were (1) Itatiaia National Park, Itatiaia

County, Rio de Janeiro State; (2) Serra dos Órgãos National Park, Teresópolis County, Rio de Janeiro State; (3) Tijuca National Park, Rio de Janeiro County, Rio de Janeiro State; (4) Parati County, Rio de Janeiro State; (5) Picinguaba State Park, Ubatuba County, São Paulo State; (6) Biological Reserve of Poco das Antas, Silva Jardim County, Rio de Janeiro State. Four collections of galls (one per season—one day in the field) were made at each of the first five of the above localities: April or May, 1998 (Autumn), July or August 1998 (Winter), October or November 1998 (Spring) and January or February 1999 (Summer). The sixth locality was sampled at monthly intervals from February 1996 to October 1997. Total numbers of each type of gall collected at each field site are given in Table 1.

Eight species of cecidomyiid flies form galls on *Mikania glomerata* in Brazil, and the galls of most of these can be distinguished on the basis of their morphology and location on the plant (Table 2) (see Gagné et al. 2001, in which gall types are also illustrated). Cecidomyiid galls were collected from *M. glomerata* plants at six localities and reared to discover what parasitoids might be attacking them. Some galls were dissected.

OBSERVATIONS

Rearings of Aspilodemon.—Rearings are summarized in Table 3. The numbers of each type of gall collected at each locality (Table 1) are estimates of their relative

Table 2. Combinity fly galls found on *Mikauia glomeratu* (Asteraceae) with descriptions of gall location and type.

Super-tribe Cecidomyiidi	
Mikaniadiplosis annulipes (Gagné)—leaf ve	in
swelling	
Tribe Clinodiplosini	
Liodiplosis cylindrica Gagné—cylindrical le	af
gall	
L. conica Gagné—conical leaf gall	
L. spherica Gagné—spherical leaf gall	
Tribe Asphondyliini	
Asphondylia glomeratae Gagné-vein swell	ing
A. moelimi Skuravá—stem swelling	
Perasphondylia mikaniae Gagné-bud gall	
Super-tribe Lasiopteridi	
Tribe Alycaulini	
Alycaulus globulus Gagné—leaf epidermis swelling	

abundance, and therefore it is clear that although the cylindrical gall type was more abundant by approximately a factor of 2 at the three localities from where most individuals of *Aspilodennon* were obtained (Parati, Picinguaba and Poço), the *Aspilodemon* demonstrated a clear preference for the spherical gall type which yielded proportionately far more wasps (Table 3).

Except for the only individual obtained from the leaf vein swelling or vein swelling (we could not differentiate these two gall types) at Picinguaba, all *Aspilodemon* individual were obtained from the galls induced by species of *Liodiplosis*. As we collected and reared many galls it is possible that the individual supposed to have been obtained from leaf vein swelling or vein swelling is a contamination.

Numerous Chalcidoidea were reared from the three gall types attacked by *Aspilodemon*; at least 10 species from the conical leaf gall type alone (taking all localities and sampling dates into account). The importance of *Aspilodemon* as a parasitoid (in terms of parasitism rate) varied: it was the commonest parasitoid reared from the spherical and conical leaf galls and the fourth commonest in the cylindrical gall.

Evidence that Aspilodemon is a parasitoid

Table 3. Total *Aspilodemon* rearings from cecidomyiid galls on *Mikania glomerata* at each site.

	Morphological type of gall							
Locality	Cylindrical	Spherical	Conical	Leaf/Vein				
Parati	- 19	65	0	0				
Itatiaia	1	0	0	0				
Ροςο	2	44	3	0				
Picinguaba	4	9	2	1				
S. Órgãos	0	2	0	0				

and not a gall-former.—Aspilodemon individuals were always found inside Cecidomyiidae gall chambers, and all the types of gall found (Table 1) yielded cecidomyiid flies showing that no gall type yielded only Aspilodemon and no potential hosts. Further, the Aspilodemon species was associated with three morphologically different gall types which would not be expected if it was a gall inducer itself.

Evidence that Aspilodemon *may be ectoparasitic.—Aspilodemon* pupates inside a cocoon which occupies approximately half of the cecidomyid chamber. All the endoparasitoids that were also reared (see above) pupated inside the cecidomyid skin and we could find the cecidomyid larval sternal spatula (a strongly sclerotised thoracic feature) within the galls after these endoparasitoids had emerged. When *Aspilodemon* parasitised the cecidomyid no host remains could be found, perhaps indicating that the sternal spatula had been consumed.

DISCUSSION

The discovery that *Aspilodemon* is associated with galls strengthens the possibility that this way of life is the norm for the subfamily Hydrangeocolinae. Previously published suggestions that *Aspilodemon* belongs to the Opiinae (Fischer 1966) have been superseded by both morphological (Wharton 1988, Whitfield 1993) and molecular phylogenetic analyses (Belshaw et al. 2000), which instead support a relationship with the Australian subfamily Mesostoinae and also with the Aphidiinae.

Several transitions to cecidogenesis have occurred in the Braconidae: Mesostoinae appear to be exclusively cecidogenic on Banksia species (Proteaceae) (Quicke and Huddleston 1989, Austin and Dangerfield 1998); the enigmatic genus Monitoriella Hedqvist produces galls on Philodeudron (Liliaceae) (Infante et al. 1995); several species of the doryctine genus Allorhogas Gahan (Macêdo and Monteiro, 1989; Marsh et al. 2000), and another doryctine, Pseuobolus, is an inquiline in figs (Ramirez and Marsh 1996) and may be partly cecidogenic. Knowledge that at least some Hydrangeocolinae (precise biology of Hydraugeocola is still unknown) are specialist parasitoids of gall-forming Diptera while the closely related Mesostoinae are cecidogenic suggests that the latter biology could have evolved from the former, as has been suggested for cecidogenic Eurytomidae (Chalcidoidea). It would be interesting to know whether any hydrangeocolines have also made this transition.

We wish to emphasise that we have not yet obtained any strong evidence about whether Aspilodemon is ecto- or endoparasitic. Although no host remains were found in galls parasitised by this wasp, indicating that all the host cuticle had been consumed, it should be noted that most endoparasitic braconids (those belonging to the 'helconoid' and 'microgastroid' groups of subfamilies—see Belshaw et al. 2000), very often emerge from their hosts before pupation and have an external feeding phase (Shaw and Huddleston 1991, Shaw and Quicke 2000). So even if the sternal spatula has been consumed it does not necessarily mean that the parasitoid was not endoparasitic in its earlier instars.

Aspilodemon differs from Hydrangeocola in only one fixed character—the fore wing pterostigma is narrow but distinct from vein R1 in the latter, whereas there is no discernible pterostigma in the former. In addition, Aspilodemon always lacks fore wing vein 2a, whereas it is present in most *Hydrangeocola*. Given that for both these characters, *Hydrangeocola* displays (at least most species) the putatively plesiomorphic state, it is likely that the species classified under *Aspilodemon* may simply be a derived group of species within a broader concept of *Hydrangeocola*.

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